

## **Allelopathic effects of *Juglans regia* L. leaf extracts on seedling growth of *Isatis tinctoria* L.**

Qian Li<sup>1,2</sup>, Zh. J. Sun<sup>1</sup>, Sh. X. Zhang\*, J.L. Liu<sup>1</sup> and K.M. Zhang<sup>3</sup>

Research Institute of Environmental Engineering,  
Ningxia University, Yinchuan, NingXia 750021, China.

E-mail: li\_q@nxu.edu.cn.

**(Received in revised form: May 07, 2019)**

### **ABSTRACT**

We studied the allelopathic effects of aqueous leaf extracts from *Juglans regia* L. on growth (seedling growth and biomass) and physiological parameters (photosynthesis, chlorophyll, soluble protein, soluble sugar, proline, malondialdehyde content) and enzymatic activities (peroxidase, superoxide dismutase and catalase enzymes) of *Isatis tinctoria*. The lowest concentration of aqueous extract (0.005 g/mL) stimulated the seedling growth and biomass accumulation. All concentrations assayed (0.005 g/mL, 0.05 g/mL and 0.1 g/mL) inhibited the respiration rate, net photosynthesis rate, stomatal conductance and chlorophyll content of *Isatis tinctoria* seedlings. However, they also increased the intercellular CO<sub>2</sub> of *Isatis tinctoria* seedlings. The contents of soluble protein, soluble sugar, proline, malondialdehyde content and the activities of the peroxidase, superoxide dismutase and catalase enzymes varied with leaf extract concentrations.

**Key word:** Allelopathic effects, aqueous extracts, *Isatis tinctoria*, *Juglans regia* L., leaf, physiological responses, seedling growth.

### **INTRODUCTION**

Allelopathic effects are widespread in agroforestry ecosystems, affecting the structure of community, succession and component crop yield (1,22,34,35). The problem of soil sickness in continuous cropping, forest regeneration and biological invasion are related to allelopathy (2,24,30,36). The studies of inter-specific relationships in agroforestry system helps us to understand the stability of ecosystem structure and function, and provides theoretical guidance to explore the rational and efficient use of resources and selection of suitable tree species for agroforestry systems (25). The mass concentration effect of allelochemicals of donor plants is an important aspect of allelopathy. The allelochemicals concentration in donor plants affects the biological activity and biomass of receptor plants (31).

The allelopathic potential of > 100 plants have been investigated, but, there are few reports on forest allelopathy (6,11,15,20,38,40). *Juglans regia* L. is major forest specie in China and has allelopathic effects on adjacent plants (9,21,27). Its major allelochemical

---

\*Correspondence author, College of Forestry, Northwest A&F University, Yangling, Shaanxi 712100, China, <sup>1</sup> Research Institute of Environmental Engineering, Ningxia University, Yinchuan, Ningxia 750021, China, <sup>2</sup> China-Arab Joint International Research Laboratory for Featured Resource and Environmental Governance in Arid Regions, Yinchuan, Ningxia 750021, China, <sup>3</sup> Co-Innovation Center for Sustainable Forestry in Southern China, College of Biology and the Environment, Nanjing Forestry University, Nanjing 210037, China.

‘juglone’ affects the seed germination, seedling growth, photosynthesis, respiration and chlorophyll content of neighbouring plants (8,14,17,37,39). Its leaves contains the phenolic acids and flavonoids (28). Owing to *Juglans regia* L. high productivity, fast growth and adaptability to management, it is most suitable tree for intercropping in agroforestry systems (29). Hence, it has been planted in Agroforestry systems in large areas in the Loess Area of Northern Wei River in Shaanxi (5). However, its allelopathic effects in intercropping with Chinese herbal medicinal plants is not known. Hence, this study aimed to evaluate the effect of aqueous leaf extracts of *Juglans regia* L. on growth and physiological responses of *Isatis tinctoria* (Medicinal plant that could be intercropped with *Juglans regia* L.).



*Isatis tinctoria* seeds

*Isatis tinctoria* seedlings

*Isatis tinctoria* pot culture

## MATERIALS AND METHODS

**Leaf aqueous extracts:** Fresh leaves of *Juglans regia* L. were collected in May 2008, from the Nursery Garden, College of Forestry at Northwest A&F University, Yangling, Shaanxi, China. (East longitude 108°4'3", north latitude 34°15'43", altitude : 460 m, annual average temperature :12.9 °C, annual average rainfall : 635.1 mm). The fresh leaves were cut into 1 cm pieces of and 10 g cut material were immersed in 100 ml-distilled water for 24 h at room temperature (20-25 °C). The aqueous extract (0.1 g/mL) was filtered through 3-layers of filter paper. For the pot experiment, the extract was further diluted to 0.005 g/mL and 0.05 g/mL.

*Isatis tinctoria* seeds were purchased from the Medicinal market of Xi'an, Shaanxi, China. The seeds were surface-sterilized with 10:1 water/bleach (commercial NaOCl) solution for 5 mins, and washed six times with distilled water. Then, seeds were sown in the field and after 15- days, the seedlings of *Isatis tinctoria* were available for transplanting in pots.

**Pot culture:** Plastic pots (20 cm dia, 27 cm depth) were filled with 10 kg loess soil (pH: 7.65, Organic matter: 4.45%, Alkali-hydrolyzable N: 116 mg·kg<sup>-1</sup>, Available P: 19.8 mg·kg<sup>-1</sup>, Available K: 64.5 mg·kg<sup>-1</sup>.) Five seedlings of *I. tinctoria* were transplanted per pot. Each pot was irrigated at 10 and 30 days after Transplanting times, the experiment duration was 40-days. Each irrigation consisted of 100 mL fresh leaf aqueous extracts concentrations of (0.005 g/mL, 0.05 g/mL and 0.1 g/mL) or water (Control) as per treatment. Treatments were

replicated thrice. The seedling height and stem girth (caliper) of *Isatis tinctoria* were measured at 10, 20, 30 and 40 day after transplanting. Forty days after transplanting, the plants were uprooted and their fresh weight, dry weight, aboveground and underground biomass of each plant were measured.

**Photosynthetic indexes and Chlorophyll :** A portable photosynthesis system (Li-6400; LI-COR Inc., Lincoln, NE, USA) was used to determine the leaf transpiration rate, net photosynthesis rate, stomatal conductance and intercellular CO<sub>2</sub> concentration of *Isatis tinctoria*. A portable chlorophyll content meter (CCM-200, OPTI-science, USA) was used to determine the chlorophyll contents of *Isatis tinctoria*.

**Physiological indexes:** Leaves of *Isatis tinctoria* were collected at 10,20,30 and 40 day after transplanting, and stored in refrigerator for one month. The frozen leaves were used to determine the soluble proteins with Coomassie blue staining (41), soluble sugars by anthrone colorimetry (41), proline by the acid ninhydrin method (34), malondialdehyde (MDA) by the thiobarbituric acid method (41), the superoxide dismutase activity (SOD) by the assay system consisting in methionine, riboflavin and NBT (12), and the peroxidase (POD) and catalase (CAT) activities by a direct spectrophotometric assay (4).

**Response Index:** The Williamson's response indices (*RI*) were used as under (32):

$$T \geq C, RI = 1 - C/T, \text{ and } T \leq C, RI = T/C - 1,$$

Where, *RI*: Allelopathic effects, C: Control data, T: Treatment data

**Statistical analysis:** All data were analyzed statistically using one-way ANOVA test in the software SPSS 16.0. Differences among means were detected with Duncan's multiple range test at 5% level.

## RESULTS AND DISCUSSION

### Seedling growth

The *Juglans regia* L. leaf extracts at different concentrations influenced the stem length, root collar diameter and seedling biomass (Fig. 1, 2 and Table 1). The *Juglans regia* L. leaf aqueous extracts at lower concentration (0.005 g/mL) stimulated the stem length of *Isatis tinctoria*. After 40 days at 0.005 g/mL concentration, the stem length reached 12.35 cm. However, higher concentration of leaf extracts (0.1 g/mL) inhibited the stem length (10.97 cm at 40<sup>th</sup> days) i.e. reduction of 11.48 % over 0.005 g/mL concentration. Lower concentration of *Juglans regia* L. leaf aqueous extracts (0.005 g/mL) were slightly stimulatory to root collar than control. In first 30 days, the root growth was slower than control, but later it grew faster and at 40<sup>th</sup> day reached 7.34 mm (0.87 mm longer than control). At 40<sup>th</sup> day, with 0.05 g/mL concentration the root collar diameter reached to 6.11 mm (94.43% of control). The fresh weight of aboveground, underground biomass and dry weight of *Isatis tinctoria* seedlings increased faster with *Juglans regia* L. leaf aqueous extract concentration of 0.005 g/mL than control. The *RI* values showed that the 0.05 g/mL extract concentration, slightly promoted the plants fresh weight, underground biomass and dry weight, but slightly inhibited the aboveground biomass. The magnitude of inhibitory

effects increased with extract concentrations. After 40 days, the 0.1 g/mL extracts were inhibitory to aboveground biomass (14.06 % Inhibition) than control. The *RI* was -0.1233 and the strength of inhibition to fresh weight decreased to -0.1084.

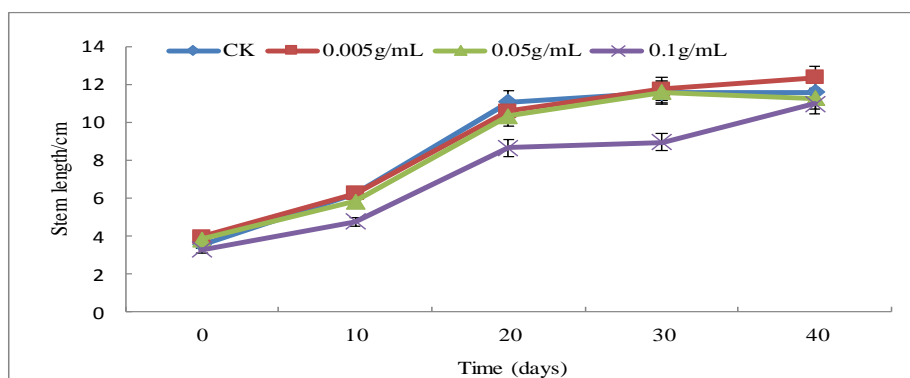


Figure 1. Effects of *J. regia* L. leaf aqueous extracts on stem length of *I. tinctoria*

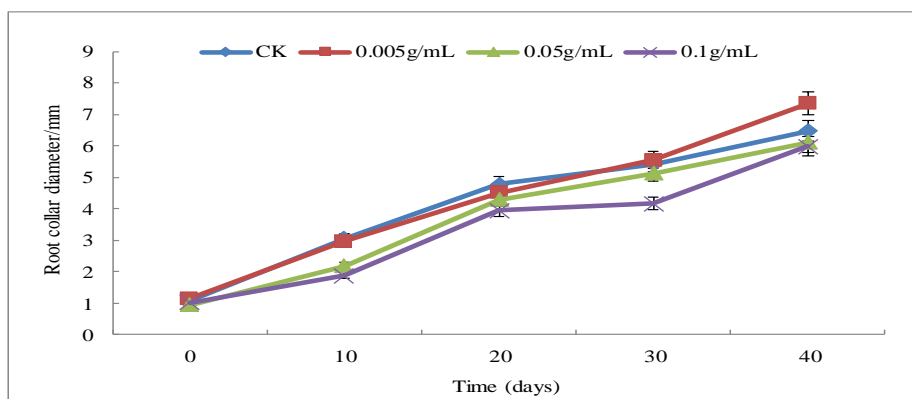


Figure 2. Effects of *J. regia* L. leaf aqueous extracts on root collar diameter of *I. tinctoria*

Table 1. Effects of *J. regia* L. leaf aqueous extracts on biomass of *I. tinctoria*

Aqueous Extract conc (g/mL)	Fresh weight/ plant		Shoot Biomass		Under ground biomass		Dry weight/ plant	
	(mg)	<i>RI</i>	(mg)	<i>RI</i>	(mg)	<i>RI</i>	(mg)	<i>RI</i>
Control	6.8374a		0.8809ab		0.4348b		1.3157bc	
0.005	8.9404a	0.2353	1.1094a	0.2060	0.5275a	0.1757	1.6306a	0.1931
0.05	7.3535a	0.0702	0.8757ab	-0.0061	0.5212a	0.1656	1.4032b	0.0624
0.1	6.0959b	-0.1084	0.7723b	-0.1233	0.3945b	-0.0927	1.1660c	-0.0032

*RI*: Response Index, Each value is mean of three replicates. Means within each column followed by the same letter are not significantly different at 5% level as per Duncan's multiple range test.

### Photosynthesis

The different concentrations of aqueous extracts from *Juglans regia* L. leaf had variable effects on the photosynthesis in *Isatis tinctoria* seedlings (Table 2). At 0.005 g/mL concentration, the seedlings respiration and net photosynthesis rates were inhibited. The respiration rate was  $2.5814 \text{ mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ , which was reduced to 36.86 % than control. The net photosynthesis rate was  $10.4722 \text{ } \mu\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ , which decreased to 35.88 % than control. The *RI*s were -0.3685 and -0.359 respectively. At 0.05 g/mL and 0.1 g/mL concentrations, the respiration rate and net photosynthesis rate were slightly inhibited. Respiration rates were  $3.2160 \text{ mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ ,  $3.6364 \text{ mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$  and the *RI*s were -0.2133 and -0.1105 respectively. Net photosynthesis rates were  $12.7945 \text{ } \mu\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$  and  $11.6667 \text{ } \mu\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ . The *RI*s were -0.217 and -0.286 separately. Three concentrations (0.005 g/mL, 0.05 g/mL and 0.1 g/mL) of aqueous extracts inhibited the stomatal conductance and chlorophyll synthesis and promoted the intercellular  $\text{CO}_2$  concentration of *Isatis tinctoria* seedlings. The *RI* of 0.005 g/mL concentrations was -0.276 and the stomatal conductance was reduced to 27.63 % than control (72.37 %). The stimulatory effects of intercellular  $\text{CO}_2$  concentration increased with the increase in extract concentrations. At 0.1 g/mL extract concentrations, the intercellular  $\text{CO}_2$  concentration reached  $303.2500 \text{ } \mu\text{L/L}$  i.e. increase of 10.03 % over control. At 0.005 g/mL, 0.05 g/mL and 0.1 g/mL extract concentrations, the chlorophyll concentration was 2.759, 2.568 and  $2.402 \text{ mg/g}$ , respectively, and the *RI*s were -0.133, -0.193 and -0.245.

Table 2. Effects of *J. regia* L. leaf aqueous extracts on photosynthesis factors and chlorophyll contents of *I. tinctoria*

Aqueous Extract conc. (g/mL)	Respiration rate ( $\text{m mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ )		Net photosynthesis rate ( $\mu\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ )		Stomatal conductance ( $\text{mol m}^{-2} \cdot \text{s}^{-1}$ )		Intercellular $\text{CO}_2$ conc ( $\mu\text{L L}^{-1}$ )		chlorophyll content (mg/g)	
	Value	<i>RI</i>	Value	<i>RI</i>	Value	<i>RI</i>	Value	<i>RI</i>	Value	<i>RI</i>
CK	4.0883a		16.3306a		246.833a		272.8333a		3.183a	
0.005	2.5814b	-0.3685	10.4722b	-0.359	178.639a	-0.276	273.9167a	0.0040	2.759ab	-0.133
0.05	3.2160ab	-0.2133	12.7945ab	-0.217	201.195a	-0.185	280.2500a	0.0264	2.568b	-0.193
0.1	3.6364a	-0.1105	11.6667ab	-0.286	199.194a	-0.193	303.2500a	0.1004	2.402b	-0.245

*RI*: Response Index, Each value is mean of three replicates. Means within each column followed by the same letter are not significantly different at 5% level as per Duncan's multiple range test.

Aqueous extracts of some plants decreased the chlorophyll content, slowed the photosynthetic rate and inhibited the seedling growth and development (7,23,36). The aqueous extracts from *Ailanthus* root inhibited the growth of *Robinia* seedling, due to decrease in chlorophyll content and biomass (3). Jia *et al.* (16) found that the mechanism of allelopathic effects in *Pinus tabuliformis-Quercus liaotungensis* mixed forest was through the soil leachates, which influenced the photosynthesis. In our experiment, the chlorophyll content of *Isatis tinctoria* seedlings decreased with increasing concentration.

### Physiological parameters

The *Juglans regia* L. leaf aqueous extracts decreased the soluble protein contents of *Isatis tinctoria* (Fig. 3). The soluble protein content decreased with treatment time. After 30 days, the inhibition rates of 0.005 g/mL, 0.05 g/mL and 0.1 g/mL concentrations decreased the soluble protein contents by 11.30 %, 12.13 % and 12.78 %, respectively, than control. The *Juglans regia* L. leaf aqueous extracts concentrations and treatment time significantly influenced the soluble sugar contents of *Isatis tinctoria* (Fig. 4). The soluble sugar content

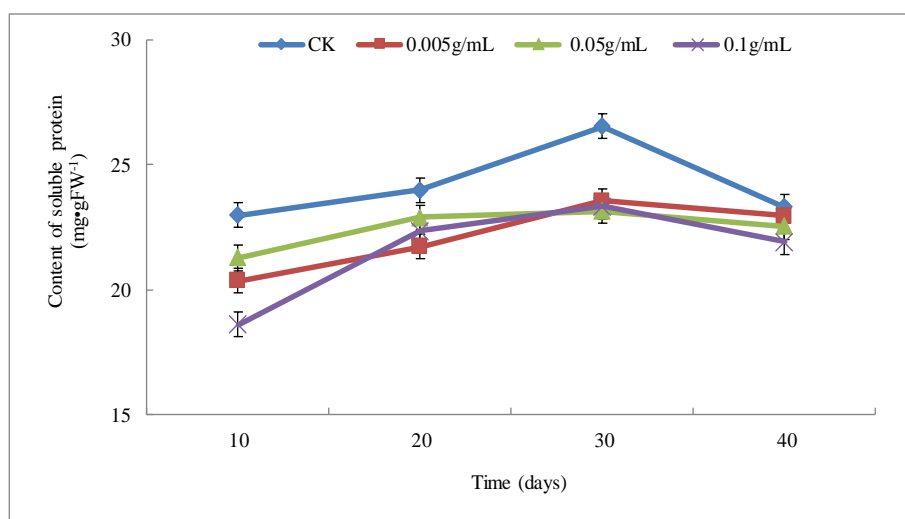


Figure 3. Effects of *J. regia* L. leaf aqueous extracts on the soluble protein content of *I. tinctoria*

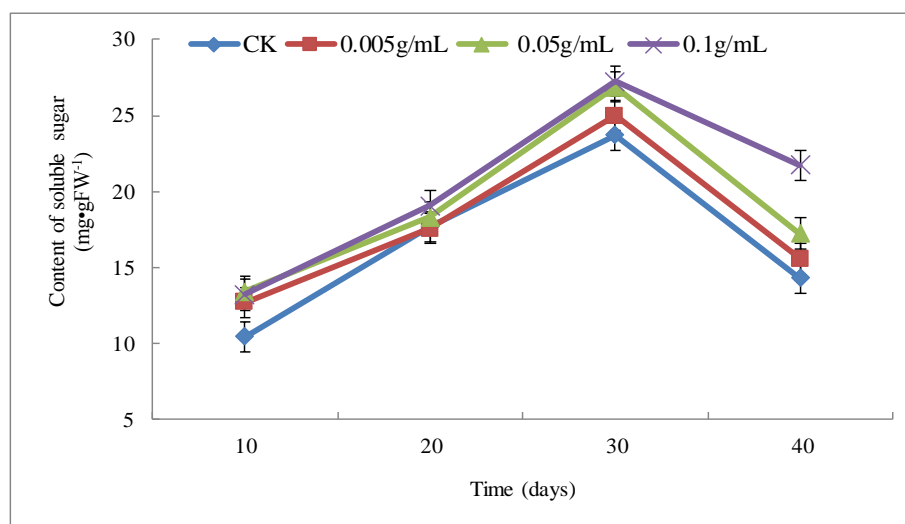


Figure 4. Effects of *J. regia* L. leaf aqueous extracts on soluble sugar content of *I. tinctoria*

of *I. tinctoria* increased with extract concentrations from 10 to 30 days and decreased thereafter. The sugar content at 30<sup>th</sup> day with 0.005 g/mL, 0.05 g/mL and 0.1 g/mL concentrations reached 24.97 mg·gFW<sup>-1</sup>, 26.87 mg·gFW<sup>-1</sup> and 27.25 mg·gFW<sup>-1</sup> respectively. The different concentrations of aqueous extracts of *Juglans regia* L. significantly influenced the PRO and MDA contents of *Isatis tinctoria* seedlings (Fig. 5, Fig. 6). After 30<sup>th</sup> days, the PRO content reached the highest and the contents in control, 0.005 g/mL, 0.05 g/mL, 0.1 g/mL were 0.236 %, 0.388 %, 0.453 % and 0.477% respectively. The MDA content increased with the increase in aqueous extracts concentration and it followed the pattern similar to PRO content i.e. increased first and then decreased. After 30 days, MDA content reached the highest and the contents in control, 0.005 g/mL, 0.05 g/mL, 0.1 g/mL and increased by 3.17 %, 50.18 % and 57.93 % respectively. These results were similar to former report showing that *Citrullus lanatus* extracts promotes the MDA content in *Lactuca sativa* leaves (10).

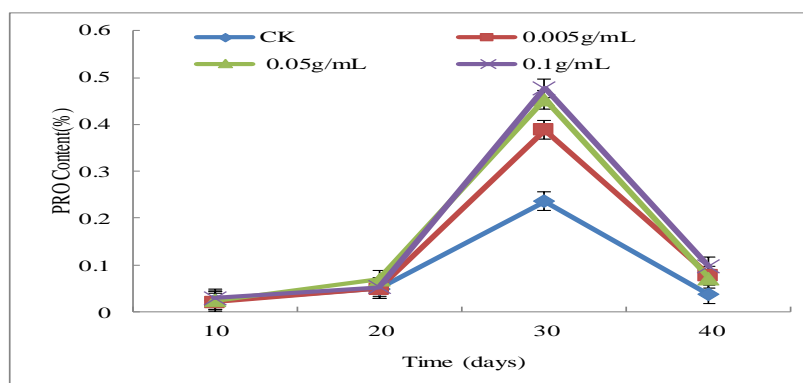


Figure 5. Effects of *J. regia* L. leaf aqueous extracts on PRO of *I. tinctoria*

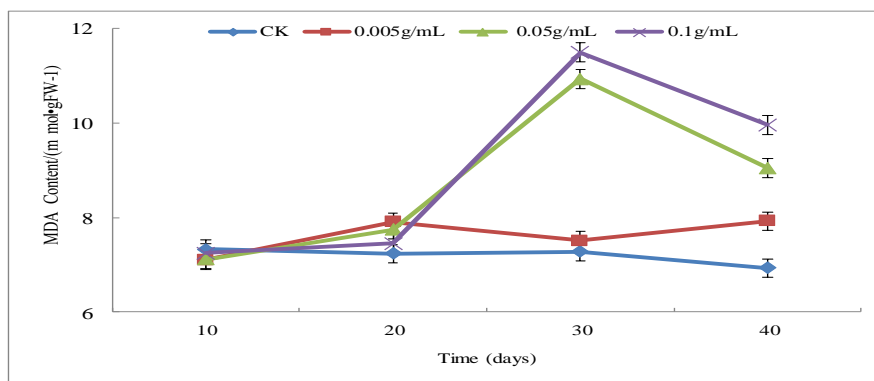


Figure 6. Effects of *J. regia* L. leaf aqueous extracts on the MDA content of *I. tinctoria*

The different concentrations of aqueous extracts of *Juglans regia* L. leaf variably influenced the POD, SOD and CAT contents of *Isatis tinctoria* (Fig. 7, Fig. 8, Fig. 9).

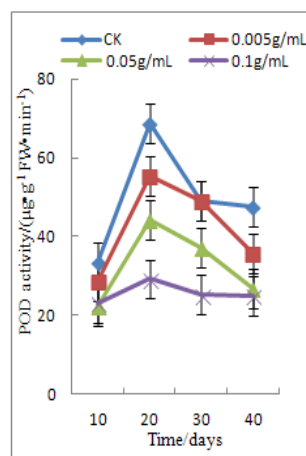


Figure 7. Effects of *J. regia* leaf aqueous extracts of POD activity of *I. tinctoria*

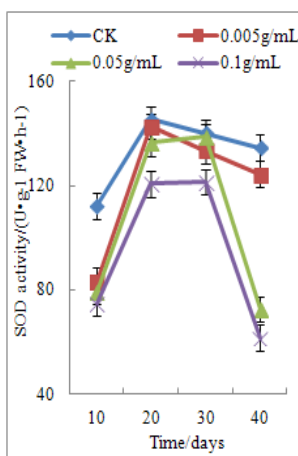


Figure 8. Effects of *J. regia* leaf aqueous extracts on SOD activity of *I. tinctoria*

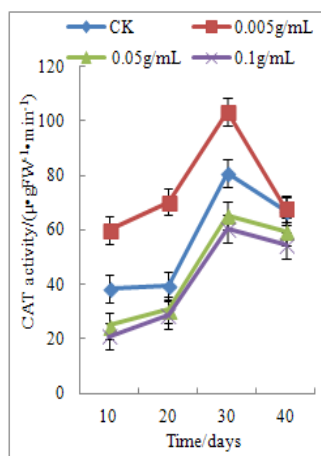


Figure 9. Effects of *J. regia* leaf aqueous extracts on the CAT activity of *I. tinctoria*

(i). **POD:** At the same concentration, the POD activity increased from the 10 to 20 days, and then decreased. After 20 days, the POD activities at 0.005 g/mL, 0.05 g/mL and 0.1 g/mL *Juglans regia* L. leaf aqueous extract reached lowest and the inhibition rates were 19.36 %, 35.72 % and 57.46 %, respectively.

(ii). **SOD:** The application of *Juglans regia* L. leaf aqueous extracts had variable effects on the SOD activity of *Isatis tinctoria* seedlings. The SOD activity decreased with the increasing concentrations. After 40 days, at 0.005 g/mL, 0.05 g/mL and 0.1 g/mL extract concentrations, the SOD activity inhibition rates were 7.55%, 46.02% and 54.19% than control, respectively. The SOD activity increased first and then decreased. At 0.005 g/mL extraction concentration, the SOD activities reached the highest on 20<sup>th</sup> day and remained constant till 30 days and decreased thereafter. The value of SOD activities were 145.38 U·g<sup>-1</sup>FW·h<sup>-1</sup> at 20<sup>th</sup> day and 142.73 U·g<sup>-1</sup>FW·h<sup>-1</sup> at 30<sup>th</sup> day with the 0.005 g/mL extraction concentration. At the end of growth period (40<sup>th</sup> day), the SOD activities in all extract concentrations decreased to normal level.

(iii). **CAT:** The application of *Juglans regia* L. leaf aqueous extracts at all concentrations increased the CAT activity till 30 days and decreased afterwards. The CAT activity reached the highest (27.84 % increase over control) on 30<sup>th</sup> day. The higher concentrations (0.05 g/mL and 0.1 g/mL) of extract treatments, inhibited the CAT activities. At 40<sup>th</sup> day, the 0.1 and 0.5 g/mL extract concentrations the inhibition rates decreased to 11.21 % and 18.90 %, respectively.

**Allelopathy Mechanism of *Juglans regia* L. leaf extracts on *Isatis tinctoria***

Many researches had shown that the aqueous extracts affects the physiological and biochemical processes in plants (26,33,39). In terms of the synthetic effect, *Juglans regia* L. extract inhibited the POD, SOD and CAT activities of *Isatis tinctoria* seedlings, which suggested that the *Juglans regia* L. leaf extracts might break the normal defence system, decrease the ability to remove H<sub>2</sub>O<sub>2</sub>, block the physiological and biochemical processes and thereby inhibit the seedling growth.

Table 3. *I. tinctoria* Synthetic analysis of allelopathic effects mechanism of *J. regia* L. leaf aqueous extracts

Function mechanism	Inhibition (%)			Synthetic effect	Allelopathic effects
	0.005 (g/mL)	0.05 (g/mL)	0.1 (g/mL)		
Soluble protein	8.51	7.19	10.99	8.90	Content decreased
Soluble sugar	-7.07	-14.69	-22.87	-14.88	Content increased
PRO	-54.51	-78.25	-89.88	-74.21	Content increased
MDA	-5.79	-21.09	-25.67	-17.52	Content increased
POD	15.23	34.56	48.46	32.75	Activity decreased
SOD	9.07	19.73	28.86	19.22	Activity decreased
CAT	-33.49	20.04	27.09	4.55	Activity decreased

The aqueous extracts from rice leaves drastically inhibit the SOD and POD activities of *Echinochloa crusgalli*, but increased the MDA content (13). Lin et al. (19) showed that autotoxins greatly influenced the cucumber root SOD and POD activities and increased the oxygen content in root tissues. The allelochemicals in aqueous extracts of *Juglans regia* L. leaf might suppress the soluble protein content, promote the soluble sugar, PRO and MDA, break the protective enzyme system and the cell membrane and increase the cell membrane permeability. The mechanism of aqueous extracts not only depended on the concentrations, but also was related to their combination and environment (18). In this study, we found that *Juglans regia* L. leaf extracts have double effects on seedling growth. The lower concentrations of *Juglans regia* L. leaf extracts had stimulatory effects and the higher concentration were inhibitory to fresh weight, shoot biomass, underground biomass and dry weight of *Isatis tinctoria*. We suggest that the changes in chlorophyll and photosynthetic factors of *Isatis tinctoria* were one of the physiological mechanisms of the allelopathic effects of *Juglans regia* L. leaf extracts. There were some water-soluble allelochemicals in *Juglans regia* L. leaf, However, the type and mass concentration of allelochemicals in *Juglans regia* L. leaf are still unclear, which requires further research.

**ACKNOWLEDGEMENTS**

This research is supported by the Major Innovation Projects for Building First-class Universities in China's Western Region (No.: ZKZD2017004) and National Natural Science Foundation of China (No.: 31660133). Many thanks are given to Mr. Zh. J. Sun, Mr. Sh. X. Zhang, Dr. F. Chen and Dr. L. Wang, for their help in field experiments.

## REFERENCES

- Bais, H.P., Vepachedu, R., Gilroy, S., Callaway, R.M. and Vivanco, J.M. (2003). Allelopathy and exotic plant invasion: from molecules and genes to species interactions. *Science* **301**: 1377-1380.
- Bais, H.P., Weir, T.L., Perry, L.G., Gilroy, S. and Vivanco, J.M. (2006). The role of root exudates in rhizosphere interactions with plants and other organisms. *Annual Review of Plant Biology* **57**: 233-266.
- Cao, B., Song, L.H. and Zhang, T. (2009). Allelopathic effects of water extracts from *Ailanthus altissima* root on growth of *Robinia pseudoacacia* seedling. *Acta Agriculture Boreali-occidentalis Sinica* **18**: 156-159. (Chinese).
- Chance, B. and Maehly, A.C. (1955). Assay of catalase and peroxidase. *Methods in Enzymology* **2**: 764-775.
- Cui, C., Cai, J. and Zhang, Sh.X. (2013). Allelopathic effects of walnut (*Juglans regia* L.) rhizospheric soil extracts on germination and seedling growth of turnip (*Brassica rapa* L.). *Allelopathy Journal* **32**: 37-48.
- Duke, S.O. (2015). Proving Allelopathy in Crop-Weed Interactions. *Weed Science* **63**: 121-132.
- Einhellig, F.A., Rasmussen, J.A. and Schon, M.K. (1979). Effects of three phenolic acids on chlorophyll content and growth of soybean and grain sorghum seedlings. *Journal of Chemical Ecology* **4**: 425-436.
- Ercisli, S., Esitken, A., Turkkal, C. and Orhan, E. (2005). The allelopathic effects of juglone and walnut leaf extracts on yield, growth, chemical and PNE compositions of strawberry cv. Fern plant. *Soil and Environment* **51**: 283-287.
- Funk, D.T., Case, P.J., Rietveld, W.J. and Phares, R.E. (1979). Effects of juglone on the growth of coniferous seedlings. *Forest Science* **25**: 452-454.
- Geng, G.D., Cheng, Z.H., Meng, H.W., Zhang, S.Q., Cao, H.X. and Zhao, H.F. (2005). Study on allelopathy and its mechanism of watermelon (*Citrullus lanatus*). *Journal of Fruit Science* **22**: 247-251.
- Ge, T.T., Huang, Y. H., He, X.L., Xu, Zh.L., Peng, Ch., Shao, H.B., Zhang D.Y. and Guo, Sh.W. (2015). Allelopathic effect of aqueous extract of different organs of sorghum on seedling growth of wheat. *Journal of Triticeae Crops* **35**: 722-728.
- Giannopolitis, C.N. and Ries, S.K. (1977). Superoxide dismutase. *I. Occurrence in higher plants. Plant Physiology* **59**: 309-314.
- He, H.Q. and Lin, W.X. (2001). Studies on allelopathic physio-biochemical characteristics of rice. *Chinese Journal of Eco-Agriculture* **9**: 56-57. (Chinese).
- Hejl, A.M., Einhellig, F.A. and Rasmussen, J.A. (1993). Effects of juglone on growth, photosynthesis and respiration. *Journal of Chemical Ecology* **19**: 559-568.
- Huang, J.B., Hu, T.X., Wu, Zh.L., Hu, H.L., Chen, H. and Wang Q. (2015). Effects of decomposing leaf litter of *Juglans regia* on growth and physiological characteristics of *Triticum aestivum*. *Acta Ecologica Sinica* **34**: 6855-6863. (Chinese).
- Jia, L.M., Zhai, M.P. and Feng, C.H. (2003). Effects of allelopathic substances on the growth and photosynthesis of *Pinus tabulaeformis* seedlings. *Journal of Beijing Forestry University* **25**: 6-10.
- Kocacaliskan, I. and Terzi, I. (2001). Allelopathic effects of walnut leaf extracts and juglone on seed germination and seedling growth. *Journal of Horticultural Science and Biotechnology* **76**: 436-440.
- Kong, C.H. and Hu F. (2001). *Allelopathy and Its Applications*. China Agricultural Press, Beijing.
- Lin, W.X., He, H.Q., Guo, Y.C., Liang, Y.Y. and Chen, F.Y. (2001). Rice allelopathy and its physiobiochemical characteristics. *Chinese Journal of Applied Ecology* **12**: 871-875. (Chinese).
- Li, W.J., Luo, K.Y., Wu D. and Luo Ch. (2017). Response of native plant species *Sophora davidii* to allelopathy of *Eupatorium adenophorum*. *Acta Ecologica Sinica* **37**: 5361-5367. (Chinese).
- MacDaniels, L.H. and Pinnow, D.L. (1976). Walnut toxicity, an unsolved problem. *North Nut Growers Association Annual Report* **67**: 114-122.
- Peng, S.L. and Shao, H. (2001). Research significance and foreground of allelopathy. *Chinese Journal of Applied Ecology* **12**: 780-786.
- Padhy, B., Patnaik, P.K. and Tripathy, A.K. (2000). Allelopathic potential of eucalyptus leaf litter leachates on germination and seedling growth of finger millet. *Allelopathic Journal* **7**: 69-78.
- Patterson, D.T. (1981). Effects of allelopathic chemicals on growth and physiological response of soybean (*Glycine max*). *Weed Science* **29**: 53-59.

25. Qin, J. and Shangguan, Z.P. (2005). Interaction effects and physiological mechanism in plants. *Agricultural Research in Arid Areas* **23**: 225-230.
26. Ren, X.Z., Guo, H.R., Ge, Y., Zhang, Y.L. and Wang, K. (2010). Allelopathy of leaf-stem and root aqueous extracts from *Potentilla acaulis* L. *Chinese Journal of Grassland* **32**: 51-56.
27. Schneiderhan, F.J. (1927). The black walnut (*Juglans nigra* L.) as cause of death of apple trees. *Phytopathology* **17**: 529-540.
28. Thevathasan, N.V., Gordon, A.M. and Voroney, R.P. (1999). Juglone (5-hydroxy-1,4 naphthoquinone) and soil nitrogen transformation interaction under a walnut plantation in southern Ontario, Canada. *Agroforestry Systems* **44**: 151-162.
29. Turk, M.A. and Tawaha, A.M. (2002). Inhibitory effects of aqueous extracts of black mustard on germination and growth of lentil. *Pakistan Agronomy Journal* **1**: 28- 30.
30. Wang, D.M., Li, D.W., Zhu, W. and Li, J.L. (2007). Allelopathic effects of different polarity allelochemicals from *Polygonatum cirrhifolium* root. *Scientia Silvae Sinica* **43**: 145-149. (Chinese).
31. Wang, P. and Zhao, X.Q. (2001). Effects of allelochemicals on cotton seed germination and seedling growth. *Journal of China Agricultural University* **6**: 26-31. (Chinese).
32. Williamson, G.B. and Richardson, D. (1988). Bioassays for allelopathy: Measuring treatment response with independent controls. *Journal of Chemical Ecology* **14**: 181-188.
33. Xu, Y.P., Tang, J.C., Gao, J.M. and Li, S.A. (2003). Allelopathic study of walnut leaf extracts. *Chemistry and Industry of Forest Products* **23**: 45-48.
34. Yan, X.F., Wang, Y. and Li, Y.M. (2007). Plant secondary metabolism and its response to environment. *Acta Ecologica Sinicae* **27**: 2554-2562. (Chinese).
35. Yu, J.Q. and Du, Y.S. (2000). Soil-sickness problem in the sustainable development for the protected production of vegetables. *Journal of Shenyang Agricultural University* **1**: 124-126.
36. Yang, L.X. (2006). Effects of *Larix gmelini* aqueous extracts on seed germination and seedling growth of *Juglans mandshurica*. *Chinese Journal of Applied Ecology* **17**: 145-1147. (Chinese).
37. Yao, H.Y., Tang, J.C. and Zhang, A.L. (2003). Advances in research of the chemistry and bioactivity of *Juglans* plant. *Acta Botanica Boreali-occidentalia Sinica* **23**: 1650-1655. (Chinese).
38. Yau, L., Cho, W.S., Yao, T., Tzi, B.N., Chi, W.T. and James, Ch.M. (2012). Research on the allelopathic potential of wheat. *Agricultural Sciences* **8**: 979-985.
39. Zhang, F.Y., Zhai, M.Z. and Mao, F.E. (2005). The effects of walnut peel's extracts on seedling growth of some crops. *Acta Agriculturae Boreali-Occidentalis Sinica* **14**: 62-65. (Chinese).
40. Zhang, Q., Yao, X.H., Teng, J.H., Shao, W.Zh. and Fu, S.L. (2016). Effects of Water Extraction from Roots of *Carya illinoensis* on seed germination and seedling growth of two crops. *Bulletin of Botanical Research* **36**: 204-210.
41. Zou, Q. (2000). *The Guide for Plant Physiological Experiments*. China Agricultural Press, Beijing.